Microbial Taxonomy and Phylogeny: Extending from rRNAs to Genomes

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Georgia Institute of Technology



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Outline

Translating old standards to sequence: The ANI approach

New insights into the species issue from natural populations

• Assessing the whole prokaryotic diversity & taxonomy

Conclusions & Perspectives



The most popular species definition

"...<u>a genomically coherent</u> (discreet) group of strains based on the hybridization of their purified DNA molecules"

...Plus, a diagnostic phenotype

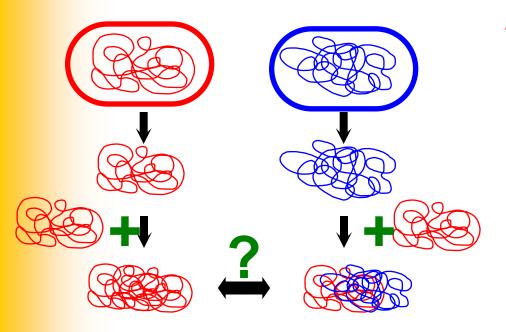


Wayne et al. IJSB, 1987

The DNA-DNA hybridization method

DDH general principle

Isolate genomic DNA from strains A and B
Random fragmentation
Denature DNA
Mix and let renature
Quantify heteroduplex relative to homoduplex



·>70% => SAME species

•Good correspondence with phenotypically coherent clusters of strains in *Enterobacteriaceae*

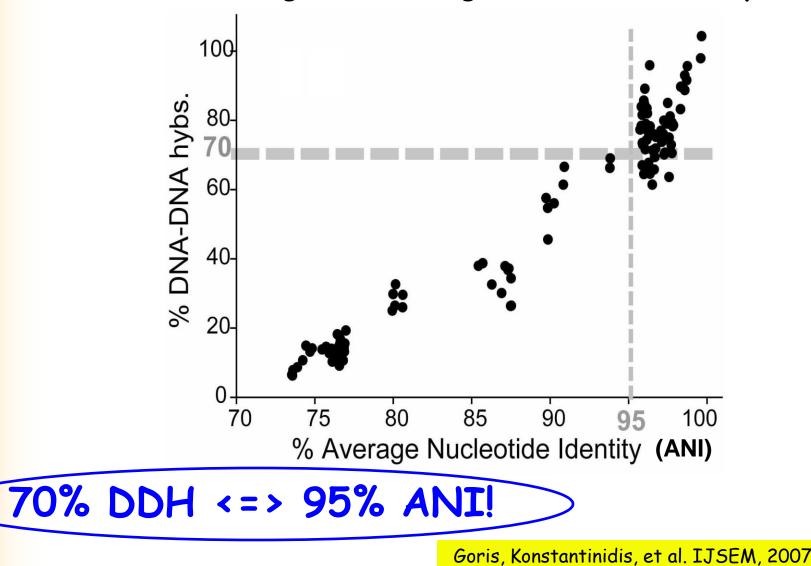
<u>BUT</u>

Difficult to do!
Unclear how it relates to whole-genome relatedness.
Need to have isolates available...but only 1-2% of prokaryotic cells are cultivable! (the great plate count anomaly)



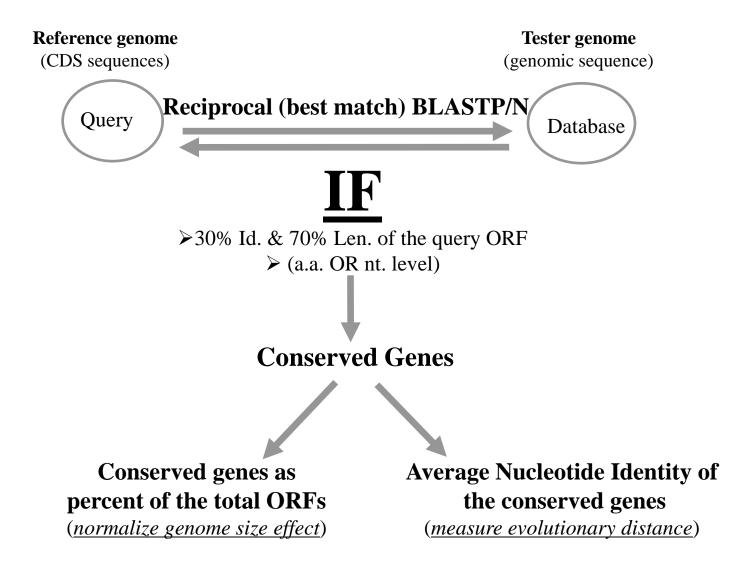
DDH vs. whole-genome sequence relatedness







ANI to measure relatedness





ANI vs. Maximum Likelihood (ML)

Within A Group

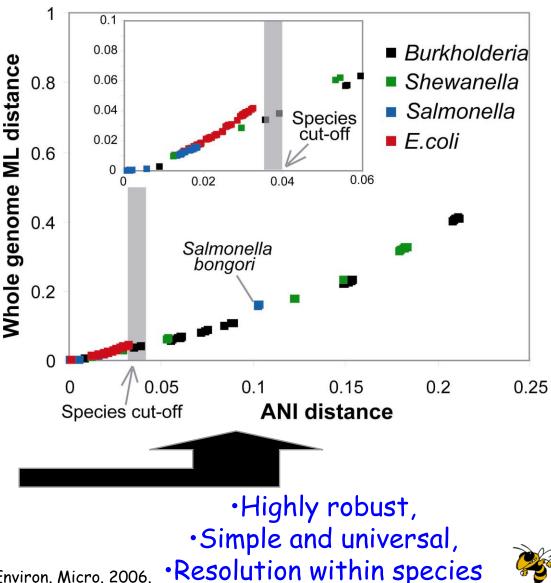
Determine the conserved gene core based on RBM blast searches.
 Build a clustalw concatenated alignment of all core genes (>2,000).

Determine the best model for sequence evolution by ModelTest.

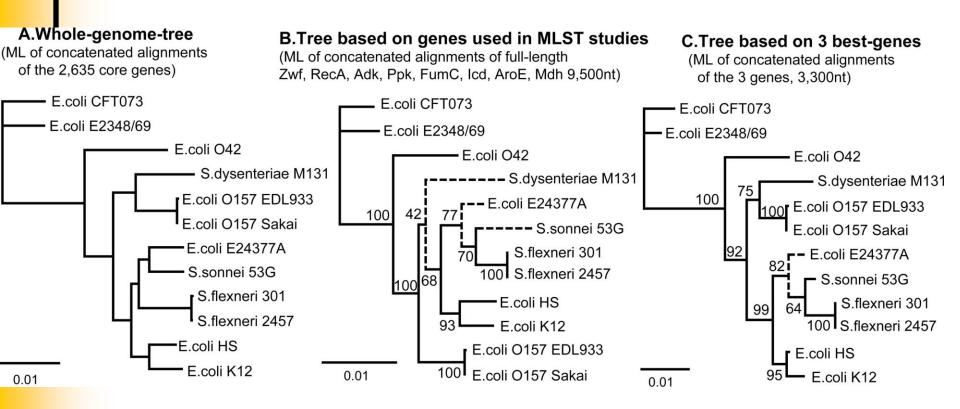
Build core-based ML phylogeny and calculate the distances between the genomes of the group.

Compare ML to ANI (calculate on the same, core genes) distances.

Konstantinidis, Ramette, &Tiedje, Applied & Environ. Micro. 2006. 👘



MLSA vs. 3-best genesin the genome



Classical MLST: Significantly different by KH test (p-value 0.02)
 Best-genes: NOT significantly different by KH test (p-value 0.709)

•Resolution at the strain level with (just) three genes!

Konstantinidis, Ramette, & Tiedje, Applied & Environ. Micro. 2006.



The most popular species definition

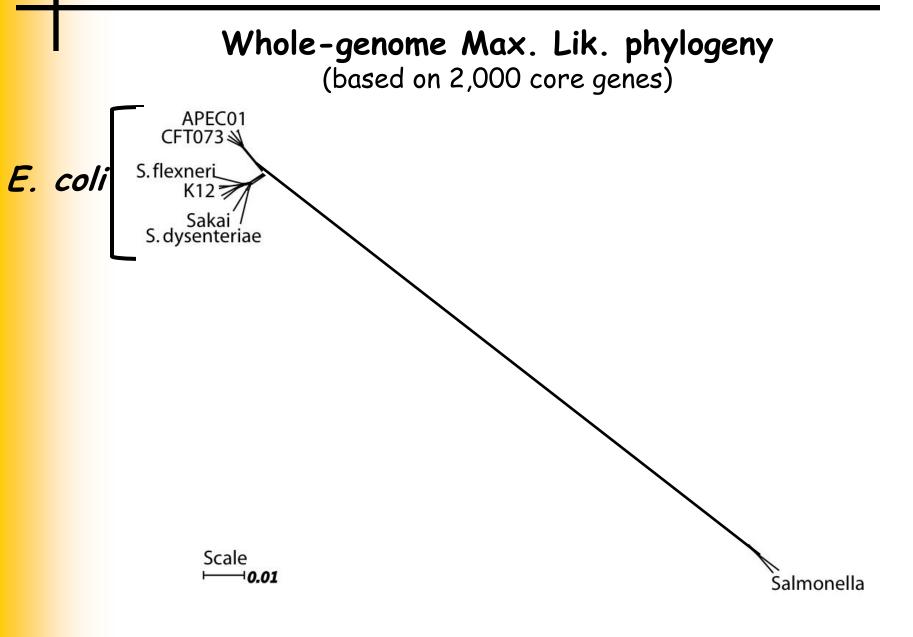
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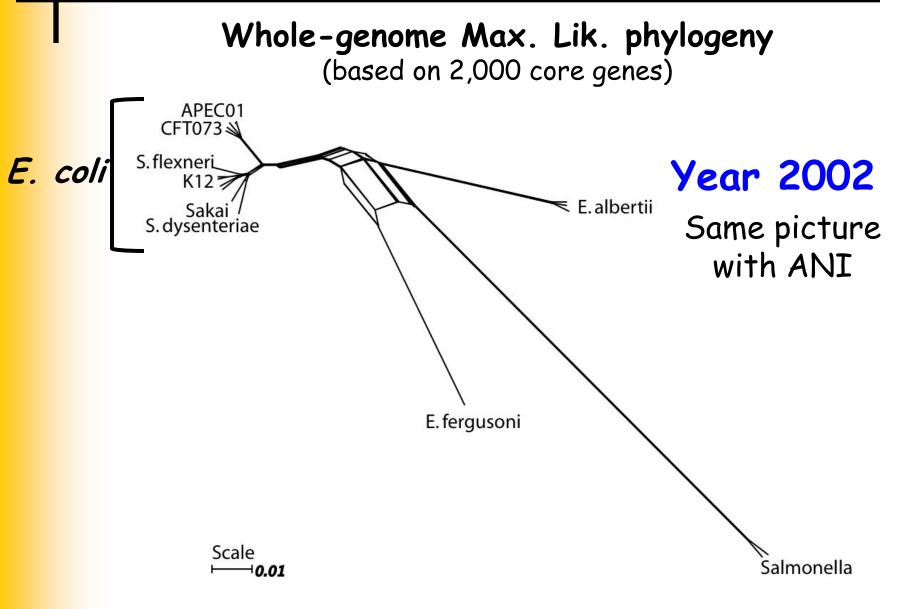
Wayne et al. IJSB, 1987

Genetic continuum OR discreet clusters?



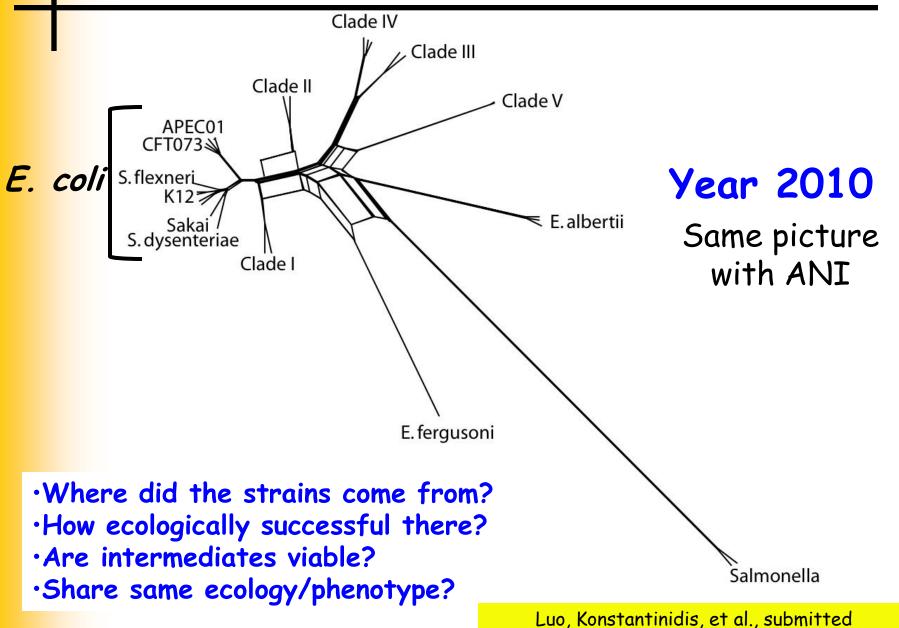


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How to study natural populations?

UNDERSTANDING OUR MICROBIAL PLANET

THE NEW SCIENCE OF METAGENOMICS



NATIONAL ACADEMY OF SCIENCES NATIONAL ACADEMY OF ENGINEERING INSTITUTE OF MEDICINE NATIONAL RESEARCH COUNCIL





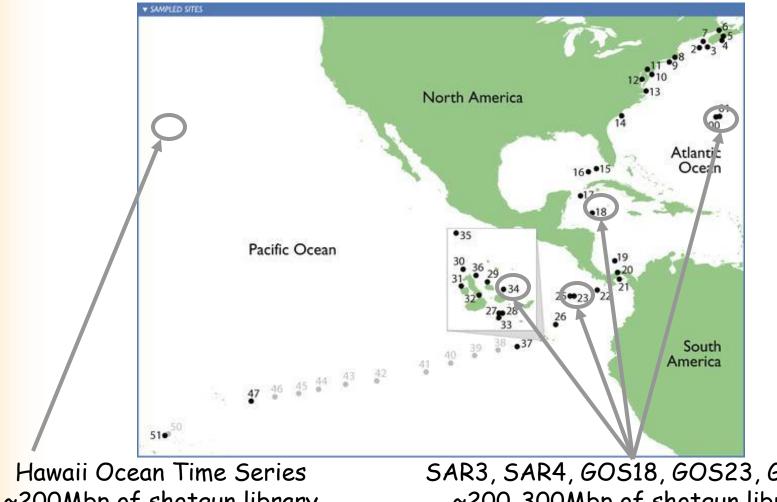
"the application of modern genomics techniques to the study of communities of microbial organisms directly in their natural environments, bypassing the need for isolation and lab cultivation of individual species"

Handelsman et al. Chemistry Biology, 1998



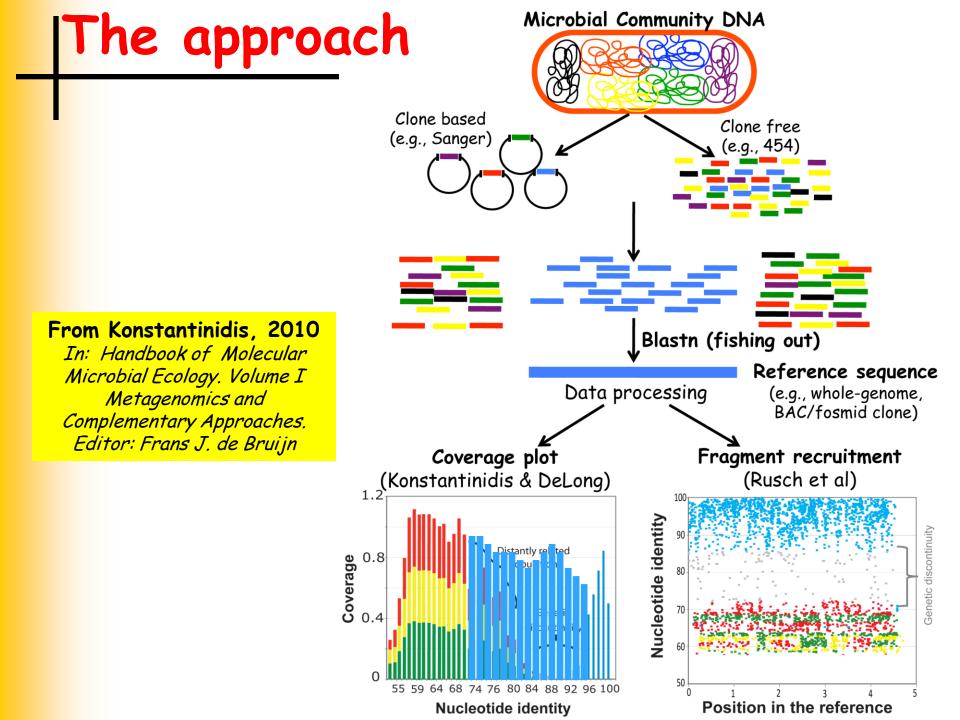
Metagenomic sampling of the Oceans

Global Ocean Survey (GOS) sampling sites

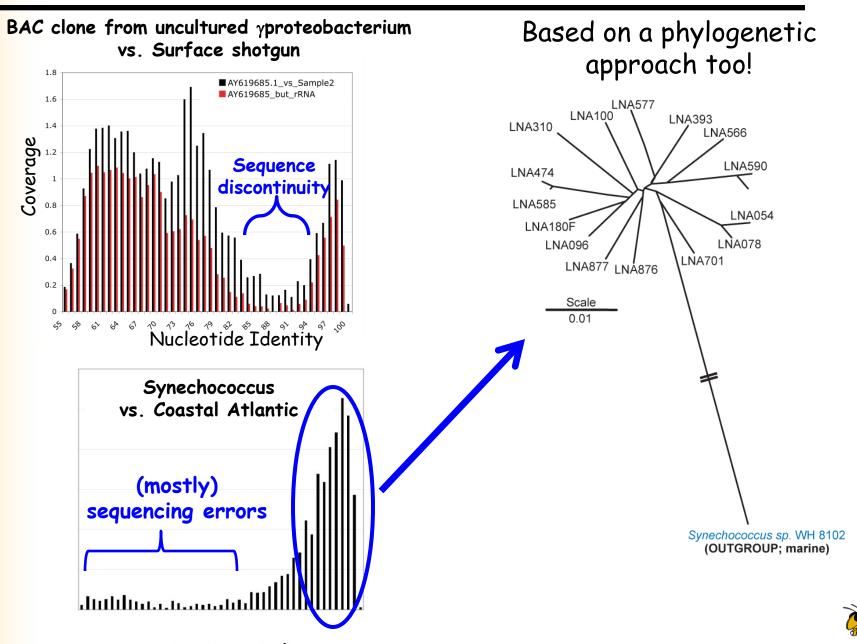


~200Mbp of shotgun library Deep, 4000m depth (Konstantinidis & DeLong, ISME 2008) SAR3, SAR4, GOS18, GOS23, GOS34 ~200-300Mbp of shotgun library Surface, ~5m depth (Rusch et al, PLoS Biology 2007)

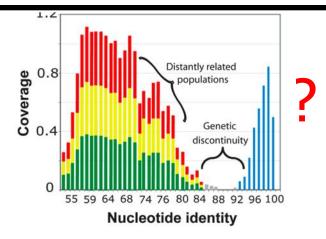




Clusters are ubiquitous in the Oceans!



Species-like populations!



- Sequence-discrete populations
- Smaller intra-population gene-content differences compared to several named species (<5% vs 20-30%)
- Detectable intra-genomic homologous recombination; albeit lower levels compared to biofilm communities (e.g., AMD)

Genetic discontinuity frequently @ 95% ANI!

More details: Konstantinidis & DeLong, The ISME Journal, 2008

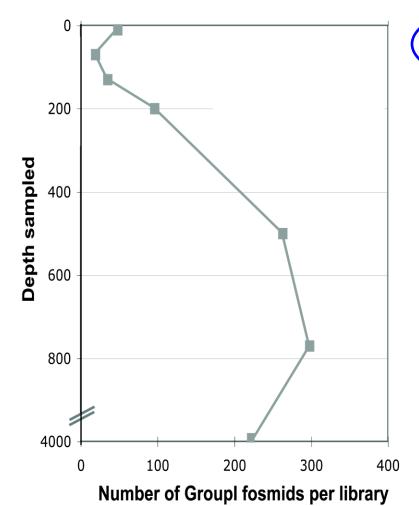


What about populations from different environments or sites?

GroupI Crenarchaea in the Oceans

Archaeal dominance in the mesopelagic

zone of the Pacific Ocean



Constituting up to 20% of microbial cells in the sub-photic zone

Genomic analysis of the uncultivated marine crenarchaeote *Cenarchaeum symbiosum*

Steven J. Hallam*[†], Konstantinos T. Konstantinidis*, Nik Putnam[‡], Christa Schleper[§], Yoh-ichi Watanabe¹, Junichi Sugahara^{||}, Christina Preston**, José de la Torre⁺⁺, Paul M. Richardson⁺, and Edward F. DeLong*⁺⁺

Hallam Konstantinidis et al. PNAS 2006

Pathways of Carbon Assimilation and Ammonia Oxidation Suggested by Environmental Genomic Analyses of Marine *Crenarchaeota*

Steven J. Hallam¹, Tracy J. Mincer¹, Christa Schleper², Christina M. Preston³, Katie Roberts⁴, Paul M. Richardson⁵, Edward F. DeLong^{1*}

Hallam et al. Plos Biol. 2006

Isolation of an autotrophic ammonia-oxidizing marine archaeon

Martin Könneke¹*†, Anne E. Bernhard¹*†, José R. de la Torre¹*, Christopher B. Walker¹, John B. Waterbury² & David A. Stahl¹

Wuchter et al. PNAS. 2006

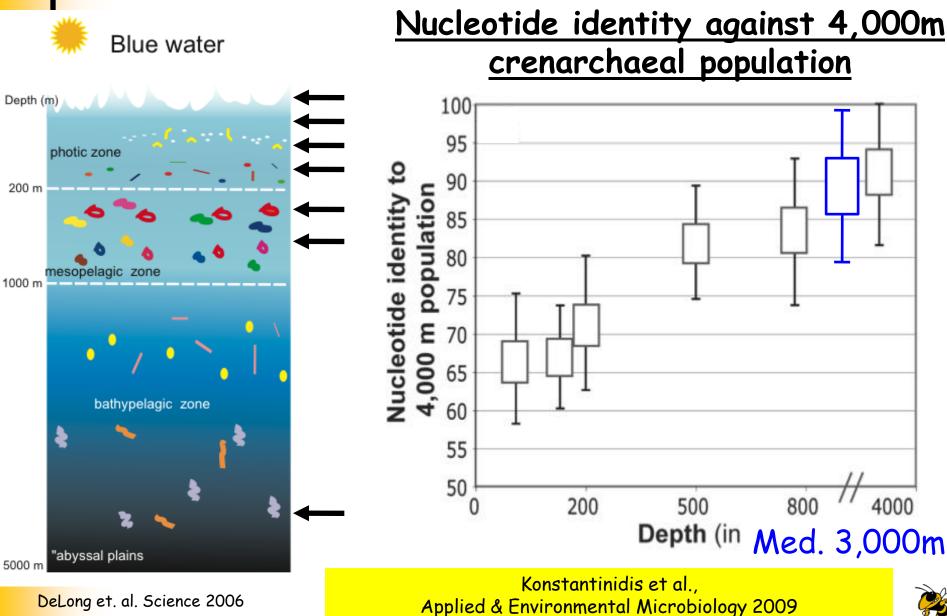
Archaeal nitrification in the ocean

Cornella Wuchter*, Ben Abbas*, Marco J. L. Coolen**, Lydie Herfort*, Judith van Bleijswijk*, Peer Timmers*, Marc Strous*, Eva Teira*5, Gerhard J. Herndi*, Jack J. Middelburg*, Stefan Schouten*, and Jaap S. Sinninghe Damsté*

Konneke et al. Nature, 2005



How are they distributed with depth?





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The current taxonomic system

•Prokaryotic Taxonomy: <u>Classification</u>, Identification, Nomenclature

•There is no official prokaryotic taxonomy. Bergey's is the closest approximation to this.

•8 recognized taxonomic ranks.

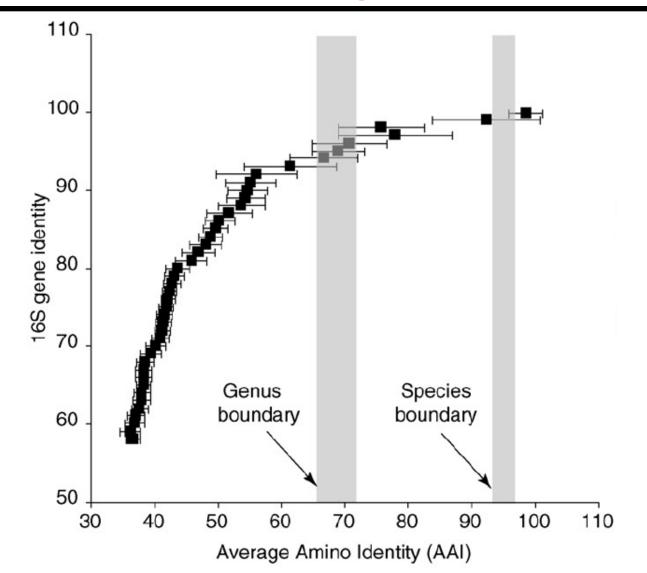
Domain Phylum Class Order Family Genus Species Subspecies

•Current classification is primarily based on 16S rRNA and secondarily on classical microscopic and biochemical observations.

How well does 16S rRNA represent the whole genome phylogeny?



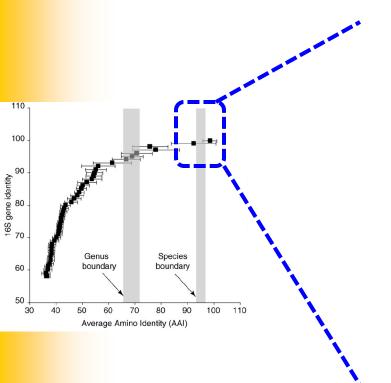
165 rRNAvs whole-genome relatedness



From Konstantinidis and Tiedje, Current Opinion in Microbiology, 2007



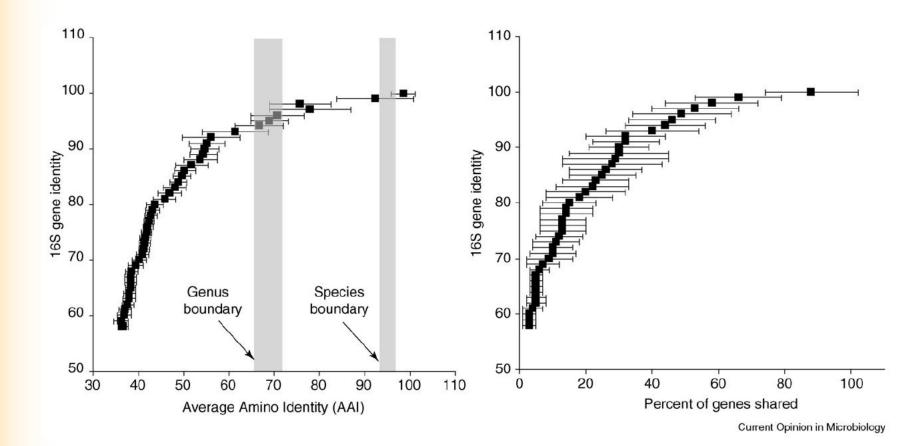
Zooming in at the species level



>70% DDH =>>95% ANI =>>98.5% 16S rRNA



165 rRNAvs gene-content relatedness



Genetic distances correspond to (even greater) gene content differences!!

From Konstantinidis and Tiedje, Current Opinion in Microbiology, 2007



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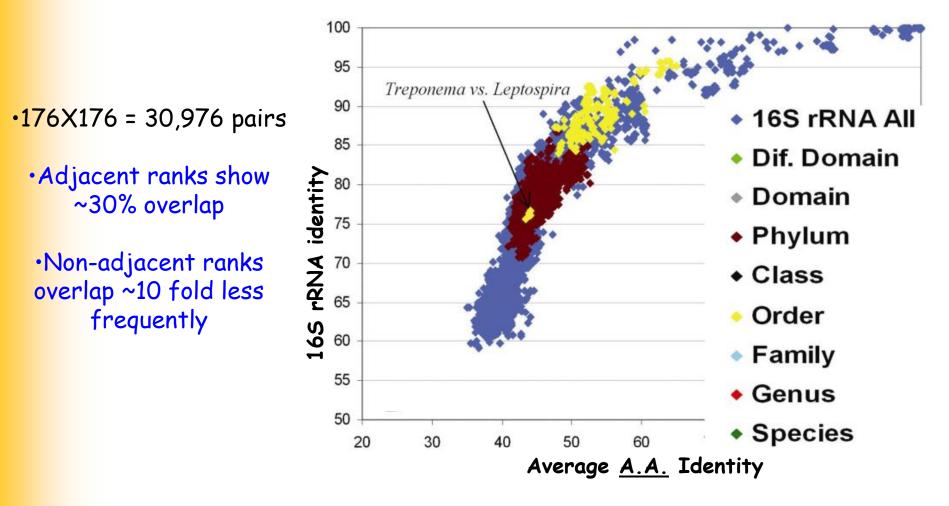
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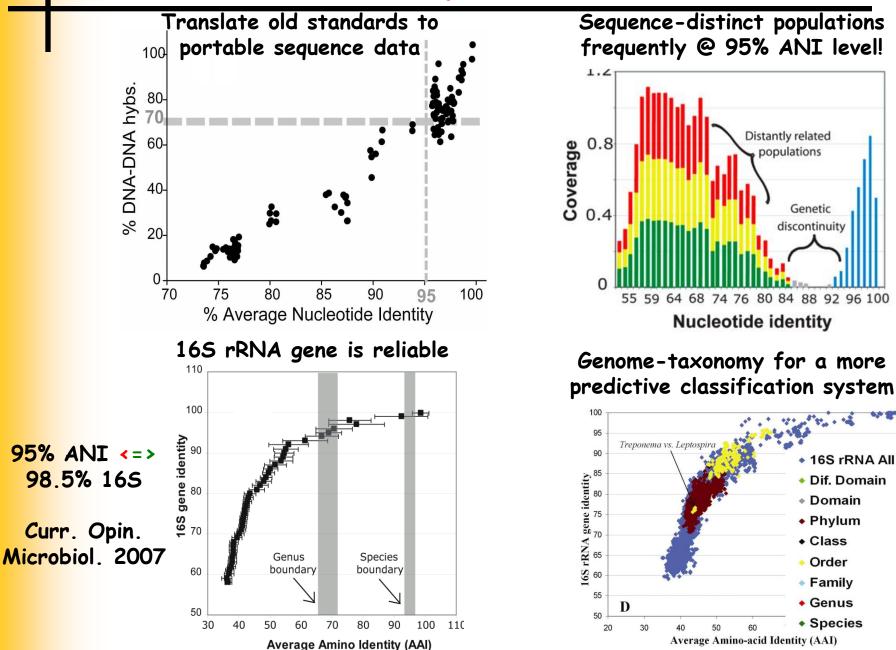
 Designation of higher than the species ranks is rather arbitrary (clustering by 165 rRNA but no standards on absolute differences).

Identifying outliers of taxonomy



From Konstantinidis and Tiedje, Journal of Bacteriology, 2005

Summary-Conclusions



Environmental Microbial Genomics Lab @ GaTech



Microbial & interdisciplinary research @ GaTech

Environmental Engineering Building Downtown Atlanta



Interested? Email kostas@ce.gatech.edu



Acknowledgments

<u>People</u>

Kostas' group @ Georgia Tech

AlejanderLuo, Bioinformatics
Alejandro Caro, Microbial Ecology/Bioinformatics
Natasha DeLeong, Microbial Ecology
Seung-Dae Oh, Env. Engineering
DespoinaTsementzi, Env. Engineering
Tate Nixon, Env. Engineering

DeLong's group @ M.I.T.

Prof. Steven Hallam (now @ UBC)
Dr. Virginia Rich (now @ U of Arizona)
Dr. Gene Tyson (now @ U of Queensland)

<u>Support</u>



Genomes to Life Program

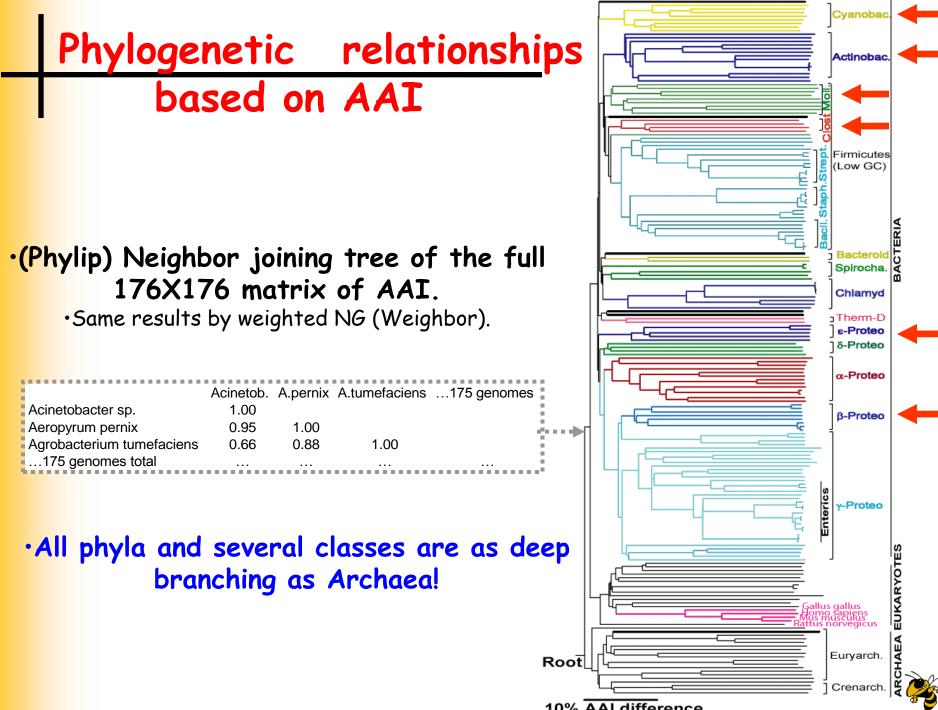
Award #DE-FG02-07ER64389



Other collaborators

Prof. Frank Loeffler (GaTech), on bioremediation communities Prof. Spyros Pavlostathis (Gatech), antibiotic resistance Prof Hang Lu (GaTech), on single-cell genomics Prof. James Tiedje (MSU), on the species concept Dr. Alban Ramette (Max Planck, Bremen), on biogeography





10% AAI difference