



Screening for Novel Biodegraders in Metagenomic Libraries of Petroleum-Associated Environments

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Bioprospection: technology versus nature

- Soil microorganisms
 - $\checkmark\,$ Source for the production of natural compounds with biological activity
 - ✓ actinobacteria, *Bacillus* spp., *Pseudomonas* spp., etc.
- Majority of biotechnological products available
 - $\checkmark\,$ Isolation and screening of microorganisms and plants
 - $\checkmark\,$ Applications : medical, industrial and agricultural areas
- High rate of "rediscovery" of new products
 - ✓ Screening of culturable microorganisms
 - $\checkmark\,$ Continuous sampling of the same environments

Trends in bioprospection

- New approaches in the search of microbial products
 - ✓ New environments
 - Endophytic microorganisms, marine, polluted and extreme environments

✓ New strategies

- New methods for selective isolation
- Cultivation-independet methods: "metagenomics"









Why Metagenomics?

 Allow the access to the metabolic diversity of uncultured microorganisms (the unseen majority)



Strategy to access and exploit the soil metagenome through the construction and screening of DNA libraries derived from soil samples or soil enrichments.

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Functional Metagenomics

Chemistry & Biology, Vol. 12, 895–904, August, 2005, ©2005 Elsevier Ltd All rights reserved. DOI 10.1016/j.chembiol.2005.05.020

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Appl Microbiol Biotechnol (2007) 74:688-698 DOI 10.1007/s00253-006-0691-0

ENVIRONMENTAL BIOTECHNOLOGY



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Novel bacterial sulfur oxygenase reductases from bioreactors treating gold-bearing concentrates

New TI Z.-W. Chen · Y.-Y. Liu · J.-F. Wu · Q. She · C.-Y. Jiang · S.-J. Liu

Si

Cloned from a Metagenomic Library

Jin-Kyu Rhee, Dae-Gyun Ahn, Yeon-Gu Kim, and Jong-Won Oh*

The ISME Journal (2009) 3, 243–251 © 2009 International Society for Microbial Ecology All rights reserved 1751-7362/09 \$32.00

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ORIGINAL ARTICLE

Functional metagenomics reveals diverse β-lactamases in a remote Alaskan soil

Heather K Allen^{1,2}, Luke A Moe¹, Jitsupang Rodbumrer^{1,3}, Andra Gaarder¹ and Jo Handelsman¹



Novel enzymes ????

Petroleum industry



Screening by PCR phenol hydroxylase large subunit gene (LmPHs)





2- BAT library - 11 positive clones

 ✓ Positive control: Acidovorax sp. isolated from the enrichment after acclimation



Functional Screening: colorimetric assay



 chromatographic assays will be performed (quantitation and identification of subproducts) New genes related to phenol degradation in metagenomic libraries derived from petroleum refinery sludge

Cynthia C. Silva, Tim Sawbridge, Helen Hayden, Maira P. Souza, Ana P.R. Torres, Vânia M.J. Santiago & Valéria M. Oliveira (*in preparation*)



MG-RAST – Metabolic profile



Cofactors, Vitamins, Prosthetic Groups, Pigments 5.09% (79) Cell Wall and Capsule 2.84% (44) Potassium metabolism 0.90% (14) Photosynthesis 0.06% (1) Miscellaneous 0.64% (10) Membrane Transport 5.09% (79) RNA Metabolism 3.22% (50) Protein Metabolism 4.51% (70) Nucleosides and Nucleotides 2.06% (32) Cell Division and Cell Cycle 2.06% (32) Motility and Chemotaxis 3.80% (59) Regulation and Cell signaling 2.51% (39) Secondary Metabolism 0.32% (5) DNA Metabolism 2.51% (39) Prophage 0.13% (2) Unclassified 3.61% (56) Virulence 8.25% (128) Macromolecular Synthesis 0.06% (1) Nitrogen Metabolism 1.23% (19) Clustering-based subsystems 15.60% (242) Respiration 2.90% (45) Stress Response 2.90% (45) Sulfur Metabolism 1.68% (26) Metabolism of Aromatic Compounds 6.96% (108) Amino Acids and Derivatives 8.96% (139) Fatty Acids and Lipids 1.48% (23) Phosphorus Metabolism 1.10% (17) Carbohydrates 9.48% (147)



 Chromatographic analyses to select the best performing clones for phenol degradation;

 Evaluation of the ability of metagenomic clones to degrade other phenol derivatives;

✓ Genetic characterization of positive fosmids able to phenol degradation (new genes or pathways?);

Metagenomic studies of microbial community in petroleum samples and investigation of its biocatalytic potential

Suzan Pantaroto de Vasconcellos & Valéria Maia de Oliveira





Ovganic Geochemistry



Organic Geochemistry

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Screening for hydrocarbon biodegraders in a metagenomic clone library derived from Brazilian petroleum reservoirs

Suzan Pantaroto de Vasconcellos^a, Célio Fernando Figueiredo Angolini^b, Isabel Natalia Sierra García^a, Bruna Martins Dellagnezze^a, Cynthia Canedo da Silva^a, Anita Jocelyne Marsaioli^b, Eugenio Vaz dos Santos Neto^c, Valéria Maia de Oliveira^{a,*}

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 Metagenomic library (aerobic + anaerobic enrichment of biodegraded oil + hexadecane as carbon source): 31,000 clones

✓ Assay for hexadecane degradation:

- microplates (colorimetric assay): 72 positive hits among 5,000 clones evaluated
- chromatographic analyses: 1 positive pool (9 clones) in 8 pools evaluated
- Pool CO8 5 clones were able to degrade hexadecane (>70%)



Fig. 4. GC-MS total ion chromatogram showing hexadecane biodegradation by the clone 2B. (a) Time 0; (b) after 28 days of incubation (98% biodegradation level).





- ✓ Diversity analysis (16S rDNA libraries)
- ✓ 6 bacterial phyla detected
 —Anaerobic enrichment (N = 91)
 —Aerobic enrichment (N = 96)

0.05



Methanohalophilus portugalensis DSM7471^T (AY290717)

Vasconcellos et al., 2010

Functional Screening: lipases and proteases

✓ Lipases: screening of 5,000 clones in plate assays with tributyrin

✓ Proteases: screening of 7,800 clones in plate assays with skin milk



Structural characterization of genes encoding hydrocarbon degradation activity in microbial metagenome derived from petroleum reservoirs Isabel Natalia Sierra Garcia & Valéria Maia de Oliveira Positive fosmid clones (HC degradation) Random shotgun library Sequencing and construction Data analysis Extract DNA DNA fragments of various sizes Sonicate AGGGATGGACGTNGAGCTOCAAGAAAGGAAAAATGGGGTCNACACCATGGGATTGGATAOGTGGGAOCAG CNCCATGAAGTGAAGGAGACTAATGAACAGAACTTCTCAAAATAGOCACTGAACTTTTACTTACAGAAAG Agarose gel electrophoresis AGCTTATGTCAGOOGGCTOGAOCTOCTAGATCAGGTATTTTATTGCAAACTATTAGAAGAAGCAAAOOGA GGCTCATTTCCTGCAGAGATGGTGAATAAAATCTTTTCTAACATTTCATCAATAAATGCCTTCCATAGTA AATTCCTATTACCTGAGCTGGAGAAACGAATGCAAGAATGGGAAACTACACCCAGAATTGGAGATATCCT GCAAAAGTTGGOGOCATTOCTTAAGATGTATGGAGAATAOGTGAAGGGATTTGATAATGCAGTGGAACTG GTTA A AAOCA TGACAGAGOGTGTTOOOCAGTTTA AATCAGTGACTGAAGAGATTCAGA A ACAGAAGATCT ATATCTACAGCAGCAAGCCATTCTAATAGTGC Purify DNA from the gel LANE 1: Sonicated H. influenzae DNA LANE 2: DNA markers Assembly and gene chracterization DNA fragments - 1.6-2.0 kb Prepare a clone library



Biodegradation evaluation of metagenomic clones



Clones (CO8)	Hexadecane
	degradation
10A	86%
1A	91%
6G	92%
3B	70%
2 B	98%

Clones (CO8)	Phenanthrene	
	degradation	
10A	49%	
1A	5%	
6G	15%	
3B	21%	
2 B	44%	\triangleright



Sequencing of shotgun library (clone 1A)



Amplicons of shotgun clones derived from fosmid 1A with *primers* M13F e M13R.



Sequencing of shotgun library (clone 1A)



Future work and Comments

Sequencing of additional clones derived from fosmid 1A is being conducted in order to achieve the complete coverage of the insert (~40 kb) responsible for hexadecane degradation (92%)

Shotgun cloning and complete sequencing of fosmid 2B, able to degrade hexadecane (98%) and phenanthrene (44%), are also being carried out

Genetic sequences responsible for oil degradation processes in Brazilian reservoirs are being for the first time identified by means of the metagenome approach

Results reinforce the huge potential of metagenomics for bioprospecting hydrocarbon degradation genes from extreme environments

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